SUPPLEMENTARY METHODS

Cytotoxicity assay

The toxicity of CDH-13 was determined with an MTT assay. Cells were seeded in 96-well plates, treated with 5 mg/mL MTT and incubated for 3 h at 37 °C. The medium was removed and 150 mL of MTT solvent was added (isopropanol containing 4 mL HCL and 0.1% Nonidet P-40). The plate was incubated for 15 min and read at 590 nm with a microplate reader.

Mouse calvarial preosteoblast isolation and osteoblast differentiation

Primary mouse osteoblasts were obtained from calvarial bones excised from two-day-old neonatal mice [1]. For osteoblast differentiation, the cells were cultured with osteogenic medium containing 10 mM β -glycerol phosphate (Sigma), 50 µg/mL ascorbate-2-phosphate (Sigma), 10⁻⁷ M dexamethasone (Sigma) and 25 ng/mL human recombinant bone morphogenetic protein 2 (R&D Systems) for 7 or 21 days.

Alkaline phosphatase (ALP) assay and Alizarine red S (ARS) staining

After 7 days of osteoblast differentiation, ALP activity was detected with 5-bromo-4-chloro-3-indolyl-

phosphate/nitro blue tetrazolium (BCIP/NBT) color development substrate (Promega, Wisconsin, USA) according to the manufacturer's instructions. For the detection of mineralization, preosteoblasts were cultured in osteogenic medium for 14-21 days and stained with ARS. Briefly, cells were fixed with 4% paraformaldehyde for 10 min and stained with 2% ARS, pH 4 (Alphachem, Middlesex, UK) for 10 min. For quantification of mineralization, ARS was removed by treatment with 10% acetic acid for 30 min, heating at 85 °C for 10 min, centrifugation at 20,000 x g for 15 min and neutralization with 10% ammonium hydroxide. The supernatants were read at 405 nm with a microplate reader.

REFERENCE

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