# *TRMT61B* rs4563180 G>C variant reduces hepatoblastoma risk: a case-control study of seven medical centers

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#### ABSTRACT

N<sup>1</sup>-methyladenosine (m<sup>1</sup>A) is an essential chemical modification of RNA. Dysregulation of RNA m<sup>1</sup>A modification and m<sup>1</sup>A-related regulators is detected in several adult tumors. Whether aberrant RNA m<sup>1</sup>A modification is involved in hepatoblast carcinogenesis has not been reported. tRNA methyltransferase 61B (TRMT61B) is responsible for mitochondrial RNA m<sup>1</sup>A modification. Some evidence has shown that genetic variants of *TRMT61B* might contribute to cancer susceptibility; however, its roles in hepatoblastoma are unknown. This study attempted to discover novel hepatoblastoma susceptibility loci. With the TaqMan method, we examined genotypes of the *TRMT61B* rs4563180 G>C polymorphism among germline DNA samples from 313 cases and 1446 controls. The association of the rs4563180 G>C polymorphism with hepatoblastoma risk was estimated based on odds ratios (ORs) and 95% confidence intervals (CIs). We found that the *TRMT61B* rs4563180 G>C polymorphism correlated significantly with a reduction in hepatoblastoma risk (GC vs. GG: adjusted OR=0.65, 95% CI=0.49-0.85, *P*=0.002; GC/CC vs. GG: adjusted OR=0.66, 95% CI=0.51-0.85, *P*=0.002).

In stratified analysis, significant associations were detected in children younger than 17 months old, girls, and subgroups with stage I+II or III+IV tumors. False-positive report probability analysis validated that children with the GC or CC genotype, particularly in those <17 months of age, had a decreased risk of hepatoblastoma. The rs4563180 G>C polymorphism also correlated with expression of *TRMT61B* and the nearby gene *PPP1CB*. We identified a high-quality biomarker measuring hepatoblastoma susceptibility, which may contribute to future screening programs.

# **INTRODUCTION**

Hepatoblastoma is a rare childhood malignancy in the liver, with an annual incidence of 0.5-1.5 per million children globally [1, 2]. Approximately 1.4 per million children in China develop this disease yearly [3]. Among all cases of pediatric tumors, approximately 1% are hepatoblastoma [2]. Most children with pediatric hepatoblastoma are diagnosed between 6 months and 3 years of age [1, 2]. Although the 5-year overall survival of hepatoblastoma is approximately 70%, improvements in clinical outcomes are still needed, especially for high-risk patients with much worse prognosis [4].

The etiology of hepatoblastoma remains largely unknown [2]. Apart from putative risk factors, such as prematurity, low birth weight, and parental smoking, several lines of evidence have substantiated that genetic factors contribute to hepatoblastoma susceptibility. First, several genetic syndromes have a greatly increased predisposition to hepatoblastoma, including Beckwith-Wiedemann syndrome (BWS), Simpson-Golabi-Behmel syndrome, hemihypertrophy, and trisomy 18 [5]. Affected children are recommended to undergo screening for hepatoblastoma and other pediatric malignancies [5]. Second, other groups and ours have identified some hepatoblastoma susceptibility loci in the myeloperoxidase [6], and CCND1 [7], xeroderma pigmentosum, complementation group C (XPC) [8], methyltransferase-like 14 (METTL14) [9], METTL3 [10], METTL1 [11], high mobility group AT-hook 2 (HMGA2) [12], tRNA (guanine-N(7)-)-methyltransferase subunit WD repeat domain 4 (WDR4) [13], YTH N6methyladenosine RNA-binding protein F1 (YTHDF1) [14], and WT1-associated protein (WTAP) [15] genes using the candidate gene method. Moreover, abundant functional susceptibility loci in pivotal genes warrant investigation in hepatoblastoma.

N<sup>1</sup>-methyladenosine (m<sup>1</sup>A) is one of the essential chemical modifications of RNA. Similar to RNA m<sup>6</sup>A modification, m<sup>1</sup>A is mediated by engaged and highly conserved enzymatic machinery involving methyltransferases (writers), demethylases (erasers), and YT521-B homology (YTH) domain-containing proteins (readers of methyl groups in RNAs). m<sup>1</sup>A is observed in tRNAs, rRNAs, mRNAs, and long noncoding RNAs

(lncRNAs), with preferential enrichment in tRNAs. m<sup>1</sup>A may affect the processing, secondary and tertiary structures, stability, translation efficiency, and biological functions of RNAs. Dysfunction of m1A-associated enzymes has been known to result in cardiovascular diseases, pulmonary diseases, Alzheimer's disease, and tumorigenesis [16, 17]. tRNA methyltransferase 6 (TRMT6) and TRMT61A are implicated in the initiation of glioma, gastrointestinal cancer, and hepatocellular carcinoma (HCC) [16, 18-22]. TRMT61B is a critical mitochondria-specific tRNA methyltransferase that is predominantly distributed in mitochondria and installs m<sup>1</sup>A at position 58 (m<sup>1</sup>A58) of tRNA (Leu (UUR), (Lys), and (Ser(UCN)), as well as 16S rRNA [23, 24]. There are very few studies regarding TRMT61B thus far. Couch et al. identified TRMT61B as a susceptibility gene in ER-negative breast cancer [25]. Ali et al. showed that TRMT61B gene variants are related to m<sup>1</sup>A/G RNA abundance in the mitochondria of various tissues and are consequently linked to many disease-promoting conditions, such as abnormal blood pressure, breast cancer, and erythrodermic psoriasis [26]. Recently, TRMT61B was found to be associated with high levels of aneuploidy. TRMT61B knockdown leads to senescence and apoptosis of melanoma cell lines [27]. The roles of TRMT61B genetic variants in hepatoblastoma have not been reported. In this study, we investigated the association of a TRMT61B singlenucleotide polymorphism (SNP) with hepatoblastoma susceptibility in a cohort of 313 cases and 1446 healthy controls.

#### **RESULTS**

#### Association study

We successfully examined the genotype of the *TRMT61B* rs4563180 G>C polymorphism in 310 children with hepatoblastoma and 1444 healthy controls among the 313 cases and 1446 controls (Table 1). While performing univariate and multivariate logistic regression analyses, we found that the rs4563180 G>C polymorphism showed protective effects against hepatoblastoma under heterogeneous conditions [adjusted odds ratio (AOR)=0.65, 95% confidence interval (CI)=0.49-0.85, P=0.002]. In other words, children harboring the GC genotype showed a 35%

Genotype	Cases (N=310)	Controls (N=1444)	P <sup>a</sup>	Crude OR (95% CI)	Р	Adjusted OR (95% CI) <sup>b</sup>	<b>P</b> <sup>b</sup>
rs4563180 (HWE=0.879)							
GG	213 (68.71)	853 (59.07)		1.00		1.00	
GC	83 (26.77)	515 (35.66)		0.65 (0.49-0.85)	0.002	0.65 (0.49-0.85)	0.002
CC	14 (4.52)	76 (5.26)		0.74 (0.41-1.33)	0.312	0.74 (0.41-1.33)	0.316
Additive			0.005	0.73 (0.58-0.91)	0.005	0.73 (0.58-0.91)	0.005
Dominant	97 (31.29)	591 (40.93)	0.002	0.66 (0.51-0.85)	0.002	0.66 (0.51-0.85)	0.002
GG/GC	296 (95.48)	1368 (94.74)		1.00		1.00	
CC	14 (4.52)	76 (5.26)	0.589	0.85 (0.48-1.53)	0.589	0.85 (0.48-1.53)	0.597

OR, odds ratio; CI, confidence interval; HWE, Hardy-Weinberg equilibrium.

 $^{a}\chi^{2}$  test for genotype distributions between hepatoblastoma patients and controls.

<sup>b</sup>Adjusted for age and gender.

lower risk of hepatoblastoma than those with the GG genotype. A significant association between the *TRMT61B* rs4563180 G>C polymorphism and reduced hepatoblastoma susceptibility was also found under additive (AOR=0.72, 95% CI=0.58-0.91, *P*=0.005) and dominant (GC/CC vs. GG: AOR=0.66, 95% CI=0.51-0.85, *P*=0.002) models (Table 1).

#### Stratified analysis

Stratified analysis revealed significant associations in children younger than 17 months old (AOR=0.62, 95% CI=0.43-0.89, P=0.010), girls (AOR=0.57, 95% CI=0.38-0.86, P=0.008), and those with hepatoblastoma in stages I+II (AOR=0.65, 95% CI=0.46-0.93, P=0.018) or III+IV (AOR=0.55, 95% CI=0.34-0.88, P=0.012) (Table 2).

#### **FPRP** analysis

We also conducted false-positive report probability (FPRP) analyses for significant findings (Table 3). The prior possibility is referred to as the possibility that the association of an SNP with a disease is genuine. Our results indicated that with a high prior possibility of 0.25, all significant associations were deserving of attention. Next, when we adopted a moderate prior possibility of 0.1, the association of the *TRMT61B* rs4563180 G>C polymorphism with hepatoblastoma susceptibility remained trustworthy in the overall analysis and among girls in stratified analysis (Table 3). These results suggest that children with the GC or CC genotype have a reduced risk of hepatoblastoma, especially girls.

#### Expression quantitative trait locus (eQTL) analysis

We explored the potential effects of the *TRMT61B* rs4563180 G>C variant on the expression of *TRMT61B* and neighboring genes. Our results indicated significantly decreased nearby *protein phosphatase 1 catalytic subunit* 

*beta* (*PPP1CB*) gene expression (P=2.3e-10) (Figure 1A) and an increase in that of the *TRMT61B* gene (P=1.2e-10) in livers with the GG genotype compared with CC genotype (Figure 1B). These results suggest that the *TRMT61B* rs4563180 G>C polymorphism may affect expression of essential genes.

#### DISCUSSION

Hepatoblastoma, especially the high-risk subtype, is rare but devastating. Genetic background plays a vital role in the initiation of hepatoblastoma. Therefore, some genetic syndromes are known to predispose patients to hepatoblastoma. For affected children, routine screening every three months is an effective strategy for early diagnosis of this disease. However, there is a lack of reliable genetic susceptibility biomarkers for large-scale screening in the community [5]. Early diagnosis and effective treatment are crucial to improving childhood cancer outcomes. It is urgent to uncover more hepatoblastoma-predisposing loci.

Large case-control studies coupled with candidate gene methodology have facilitated discovery of numerous disease susceptibility genes in various disorders. However, such studies in hepatoblastoma are minimal due to its low incidence. Most case-control studies contain fewer than 100 samples from hepatoblastoma patients [6, 7]. Fortunately, we have established a relatively large cohort of 313 cases and 1446 controls over the past years. Our group has previously reported that many genetic variants of genes encoding RNA m<sup>6</sup>A and m<sup>7</sup>G methyltransferase, demethylase, and m<sup>6</sup>A-reading proteins confer hepatoblastoma susceptibility, including *METTL3* [10], *METTL4* [9], *AlkB homolog 5* (*ALKBH5*) [28], *FTO* [29], *YTHDC1* [30], *YTHDF1* [14], *WTAP* [15], *WDR4* [13], and *METTL1* [11].

m<sup>1</sup>A is also one of the essential reversible chemical modifications of RNA and is installed on the first

Variables	GG	GC/CC	Crude OR	D	Adjusted OR <sup>a</sup>	<b>p</b> a
variables	(Cases/C	Controls)	(95% CI)	- P	(95% CI)	- <i>P</i> "
Age, month						
<17	115/369	52/272	0.61 (0.43-0.88)	0.008	0.62 (0.43-0.89)	0.010
≥17	98/484	45/319	0.70 (0.48-1.02)	0.063	0.70 (0.48-1.02)	0.066
Gender						
Females	91/347	37/248	0.57 (0.38-0.86)	0.008	0.57 (0.38-0.86)	0.008
Males	122/506	60/343	0.73 (0.52-1.02)	0.063	0.73 (0.52-1.02)	0.063
Clinical stage						
I+II	110/853	50/591	0.66 (0.46-0.93)	0.018	0.65 (0.46-0.93)	0.018
III+IV	66/853	25/591	0.55 (0.34-0.88)	0.012	0.55 (0.34-0.88)	0.012

Table 2. Stratification analysis for the association between *TRMT61B* rs4563180 genotypes and hepatoblastoma risk.

OR, odds ratio; CI, confidence interval.

<sup>a</sup>Adjusted for age and gender, omitting the corresponding stratify factor.

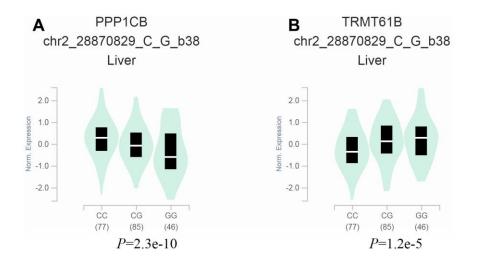
Table 3. False-positive report probability analysis for significant findings derived from *TRMT61B* rs4563180 and hepatoblastoma risk.

Comotomo	OR (95% CI)	P <sup>a</sup>	Statistical	Prior probability				
Genotype			power <sup>b</sup>	0.25	0.1	0.01	0.001	0.0001
GC vs. GG	0.65 (0.49-0.85)	0.002	0.411	0.014	0.040	0.314	0.822	0.979
GC/CC vs. GG	0.66 (0.51-0.85)	0.002	0.450	0.011	0.033	0.272	0.791	0.974
<17 months	0.61 (0.43-0.88)	0.008	0.326	0.072	0.188	0.719	0.963	0.996
Female	0.57 (0.38-0.86)	0.008	0.230	0.092	0.234	0.770	0.971	0.997
Stage I+II	0.66 (0.46-0.93)	0.018	0.459	0.107	0.265	0.799	0.976	0.998
Stage III+IV	0.55 (0.34-0.88)	0.012	0.210	0.148	0.343	0.852	0.983	0.998

OR, odds ratio; CI, confidence interval.

<sup>a</sup>Chi-square test was used to calculate the genotype frequency distributions.

<sup>b</sup>Statistical power was calculated using the number of observations in each subgroup and the corresponding ORs and *P* values in this table.



**Figure 1. Relationship between the** *TRMT61B* **rs4563180 G>C polymorphism and gene expression.** (A, B) Gene Tissue Expression (GTEx) analysis indicated downregulation of the nearby protein phosphatase 1 catalytic subunit beta (*PPP1CB*) gene and upregulation of the *TRMT61B* gene in liver tissues with the GG genotype compared to the CC genotype.

nitrogen atom of adenosine [17]. m<sup>1</sup>A modifications are much less prevalent in mRNA than m<sup>6</sup>A but preferentially occur in tRNA. Many studies have implicated RNA m<sup>1</sup>A in cardiovascular diseases, pulmonary diseases, Alzheimer's disease, and tumorigenesis [16, 17]. In HCC, m<sup>1</sup>A modification levels in tRNA are remarkably upregulated, which was attributed to the highly expressed m1A methyltransferase complex TRMT6/TRMT61A [22]. TRMT6/TRMT61A promotes PPAR<sub>δ</sub> translation by specifically enhancing m<sup>1</sup>A methylation in relevant tRNAs, further activating hedgehog signaling and stimulating self-renewal of liver CSCs and tumorigenesis [22]. Ye et al. confirmed TRMT6 overexpression in HCC tissues, and ectopic expression of TRMT6 accelerates HCC cell proliferation and stimulates cell cycle progression by activating the PI3K/AKT pathway [19]. TRMT6/61A overexpression was also observed in urothelial carcinoma of the bladder, along with dysregulation of the tRF targetome. TRMT6/61A-induced upregulated m<sup>1</sup>A modification on tRFs plays an essential role in maintaining a proper unfolded protein response [18].

TRMT61B is a crucial tRNA methyltransferase in mitochondria, which has a genome composed of 2 rRNA genes, 22 tRNA genes, and 13 mRNA genes. m<sup>1</sup>A RNA modification levels affect mitochondrial transcript processing. Genetic variants of TRMT61B have been shown to associate with mitochondrial m<sup>1</sup>A RNA modification degrees at functionally crucial sites in a broad spectrum of tissues [26]. These results implied that TRMT61B variants might have a profound impact on the function of mitochondrial RNAs. Coincidently, Couch et al. determined that TRMT61B at 2p23.2 is a susceptibility gene in ERnegative breast cancer based on functional and eQTL studies [25]. Recently, TRMT61B was found to be associated with high levels of aneuploidy, and its knockdown led to senescence and apoptosis of melanoma cell lines [27].

Given the importance of m<sup>1</sup>A in tumorigenesis and the impacts of *TRMT61B* gene variants on m<sup>1</sup>A RNA levels, it is necessary to explore the association between *TRMT61B* gene SNPs and hepatoblastoma susceptibility. In this study, we found that the *TRMT61B* rs4563180 G>C polymorphism was significantly associated with a decreased risk of hepatoblastoma. Significant associations were found for children <17 months of age, girls, and subgroups with stage I+II or III+IV disease in stratified analysis. It should be noted that genetic association studies generally require many statistical tests. In such cases, significant associations between genetic variants and susceptibility to diseases based on *P* values may only be spurious findings. Therefore, PFRP analyses were

performed to validate our significant findings by integrating the prior possibility, study power, and Pvalues. With a moderate prior possibility of 0.1, PFRP analyses indicated that the association of the TRMT61B rs4563180 G>C variant with the risk of hepatoblastoma was trustworthy in the whole population and in girls. Interestingly, GTEx analyses revealed that the TRMT61B rs4563180 G>C variant might modify expression of the PPP1CB and TRMT61B genes. The protein product of the PPP1CB gene is one of the three catalytic subunits of protein phosphatase 1 (PP1), which is a serine/threonine protein phosphatase regulating many fundamental cellular processes (e.g., cell division and protein synthesis) [31]. Recently, a genetic variant of PPP1CB was reported to modify the risk of hepatitis B virusrelated HCC [32]. Collectively, we identified TRMT61B as a high-quality hepatoblastoma gene, and the roles of its genetic variant in disease susceptibility deserve further attention.

Attention should also be given to the limitations of this study. First, we made great efforts to recruit subjects from seven independent medical centers across China. However, the sample size was moderate because of the disease's rareness, which limited the study's statistical power, especially with regard to stratified analysis. Second, the study population was restricted to Han Chinese ethnicity. Third, potential environmental risk factors were not included in analyses because of the inaccessibility of the information. Fourth, we could not perform survival analysis without relevant information. Finally, only one SNP that met the selection criteria was investigated in this study.

In summary, we discovered that the *TRMT61B* rs4563180 G>C variant significantly reduced the risk of childhood hepatoblastoma. This SNP may be used to screen for children at high risk of hepatoblastoma when combined with other essential susceptibility loci.

#### **MATERIALS AND METHODS**

#### Study population

Complete information on the cohort was provided in previous publications [15]. In brief, 313 children with hepatoblastoma and 1446 healthy controls (Supplementary Table 1) of Han Chinese nationality were recruited from participating medical centers located in different cities across China, including Taiyuan, Guangzhou, Kunming, Zhengzhou, Changsha, Xi'an, and Shenyang. All patients were newly diagnosed and confirmed by two or more pathologists. The clinical stages of the patients were determined using the PRETEXT classification [33]. Healthy volunteers were recruited from the same seven medical centers above who visited for routine examinations. Epidemiological and clinical data on these children were reported previously [15]. We obtained signed informed consent for every subject from their parents or guardians.

#### Identification and genotyping of SNPs

Following standard criteria [34, 35], we determined the rs4563180 G>C polymorphism as a potential functional SNP in the TRMT61B gene. We predicted SNP functions using the online tool SNPinfo (https://snpinfo.niehs.nih.gov/snpinfo/snpfunc.html). The results retrieved showed that the rs4563180 polymorphism is located in the transcription factorbinding site (TFBS). Therefore, this SNP may affect binding of transcription factors to the TRMT61B gene, consequently modulating its transcription. DNA was extracted from peripheral blood samples of the participants with Tiangen Blood DNA Extraction Kit (Taigen Biotechnology, Beijing, China). Genotypes of samples were examined using the TaqMan instrument (Applied Biosystems, Foster City, CA, USA) and a 384well plate format. Positive and negative control samples were run in parallel with the samples in each assay to ensure genotyping quality. Laboratory workers who performed the experiments were blinded to the identities of the specimens. Routinely, a fixed fraction of DNA samples was randomly selected for validation testing. A concordance rate of 100% for the repeated tests was needed.

#### Genotype-tissue expression (GTEx) analysis

The GTEx project was initiated by the National Institutes of Health in September 2010. This project generated a database that allows researchers to study relationship between inherited gene changes and common diseases. This resource contains expression of genes in various tissues from many different people, accompanied by genotypes of numerous SNPs [36]. This web tool was used to explore the impacts of the rs4563180 G>C polymorphism on gene expression.

#### False-positive report probability analysis

Genetic association studies are usually subjected to many statistical tests, with statistical significance determined by P values. As a result, some associations between genetic variants and diseases based on a P value below 0.05 alone may be falsely positive. To eliminate false significant associations between genetic variants and hepatoblastoma, FPRP analysis was adopted by considering the prior probability of a genuine association of the SNPs with a disorder, the power of statistics, and *P* values. Prior probabilities of 0.1, 0.01, or 0.001 were defined as high, moderate, or low, respectively. A given association with an FPRP value below 0.2 deserves attention or trustworthiness [37, 38].

#### Statistical analysis

A t test or  $\chi^2$  test was applied to check the significant differences between the cases and controls for continuous or categorical variables, respectively. A goodness-of-fit  $\chi^2$  test was employed to evaluate the Hardy-Weinberg equilibrium (HWE) of the tested SNP in the controls. Finally, the statistical significance of the association of the *TRMT61B* SNP and hepatoblastoma susceptibility was determined using unconditional logistic regression analysis. The strength of the association between the *TRMT61B* SNP and hepatoblastoma susceptibility was measured by ORs and 95% CIs. Stratified analyses by age, sex, and clinical stage were carried out. A two-sided *P*<0.05 was accepted as statistically significant. All analyses were completed with SAS v9.4 (SAS Institute Inc., Cary, NC, USA).

#### Data availability statement

All the data are available upon request.

## Abbreviations

**BWS**: Beckwith-Wiedemann syndrome; XPC: xeroderma pigmentosum, complementation group C; METTL14: methyltransferase like 14; HMGA2: high mobility group AT-hook 2; WDR4: WD repeat domain 4; YTHDF1: YTH N6-methyladenosine RNA-binding protein F1; WTAP: WT1-associated protein; m<sup>1</sup>A: N<sup>1</sup>methyladenosine; lncRNA: long noncoding RNA; TRMT6: tRNA methyltransferase 6: HCC: hepatocellular carcinoma; SNP: single-nucleotide polymorphism; OR: odds ratio; CI: confidence interval; FPRP: false-positive report probability; eQTL: expression quantitative trait locus; PPP1CB: protein phosphatase 1 catalytic subunit beta; ALKBH5: AlkB homolog 5; PP1: protein phosphatase 1; TFBS: transcription factor-binding site; GTEx: Genotype-Tissue Expression; HWE: Hardy-Weinberg equilibrium.

# **AUTHOR CONTRIBUTIONS**

All authors contributed significantly to this work. FL, LL, ZY, YL, JZhang, JC, SL, and JH performed the research study and collected the samples and data. JZhu and JH analyzed the data. DZ, JL and JH designed the research study. DZ, JZhu, JL and JH wrote the paper. JZhu and JH prepared all the tables and figures. All authors have read and approved the final manuscript to be published.

#### **CONFLICTS OF INTEREST**

The authors have no conflicts of interest to declare.

#### ETHICAL STATEMENT AND CONSENT

The study protocol was assessed and approved by the institutional review board of Guangzhou Women and Children's Medical Center (Ethical Approval No: 202016601). We obtained signed informed consent for every subject from their parents or guardians.

#### FUNDING

This study was supported by grants from the Guangxi Science and Technology Plan Project (Guangxi Clinical Research Center for Obstetrics and Gynecology, GuiKe: AD22035223) and Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease (No: 2019B030301004).

#### **REFERENCES**

- Rougemont AL, McLin VA, Toso C, Wildhaber BE. Adult hepatoblastoma: learning from children. J Hepatol. 2012; 56:1392–403. <u>https://doi.org/10.1016/j.jhep.2011.10.028</u> PMID:<u>22326463</u>
- Spector LG, Birch J. The epidemiology of hepatoblastoma. Pediatr Blood Cancer. 2012; 59:776–9. <u>https://doi.org/10.1002/pbc.24215</u> PMID:22692949
- Bao PP, Li K, Wu CX, Huang ZZ, Wang CF, Xiang YM, Peng P, Gong YM, Xiao XM, Zheng Y. [Recent incidences and trends of childhood malignant solid tumors in Shanghai, 2002-2010]. Zhonghua Er Ke Za Zhi. 2013; 51:288–94. PMID:23927803
- Meyers RL, Maibach R, Hiyama E, Häberle B, Krailo M, Rangaswami A, Aronson DC, Malogolowkin MH, Perilongo G, von Schweinitz D, Ansari M, Lopez-Terrada D, Tanaka Y, et al. Risk-stratified staging in paediatric hepatoblastoma: a unified analysis from the Children's Hepatic tumors International Collaboration. Lancet Oncol. 2017; 18:122–31. https://doi.org/10.1016/S1470-2045(16)30598-8 PMID:27884679
- Kalish JM, Doros L, Helman LJ, Hennekam RC, Kuiper RP, Maas SM, Maher ER, Nichols KE, Plon SE, Porter CC, Rednam S, Schultz KA, States LJ, et al. Surveillance Recommendations for Children with Overgrowth Syndromes and Predisposition to Wilms Tumors and Hepatoblastoma. Clin Cancer Res. 2017; 23:e115–22. https://doi.org/10.1158/1078-0432.CCR-17-0710

PMID:28674120

 Pakakasama S, Chen TT, Frawley W, Muller C, Douglass EC, Tomlinson GE. Myeloperoxidase promotor polymorphism and risk of hepatoblastoma. Int J Cancer. 2003; 106:205–7. <u>https://doi.org/10.1002/ijc.11191</u>

PMID:<u>12800195</u>

- Pakakasama S, Chen TT, Frawley W, Muller CY, Douglass EC, Lee R, Pollock BH, Tomlinson GE. CCND1 polymorphism and age of onset of hepatoblastoma. Oncogene. 2004; 23:4789–92. <u>https://doi.org/10.1038/sj.onc.1207499</u> PMID:<u>15133490</u>
- Zhuo Z, Miao L, Hua W, Chen H, Yang Z, Li Y, Zhang J, Li S, Cheng J, Li L, Xia H, He J. Genetic variations in nucleotide excision repair pathway genes and hepatoblastoma susceptibility. Int J Cancer. 2021; 149:1649–58. <u>https://doi.org/10.1002/ijc.33722</u> PMID:34196959
- Chen H, Chen Z, Wang M, Zhang J, Li Y, Li L, Li S, Cheng J, Wang X, Xia H, Yang Z, He J. METTL14 gene polymorphisms influence hepatoblastoma predisposition in Chinese children: Evidences from a seven-center case-control study. Gene. 2022; 809:146050. https://doi.org/10.1016/j.gene.2021.146050

PMID:34743822

- Chen H, Duan F, Wang M, Zhu J, Zhang J, Cheng J, Li L, Li S, Li Y, Yang Z, Xia H, Niu H, He J. Polymorphisms in METTL3 gene and hepatoblastoma risk in Chinese children: A seven-center case-control study. Gene. 2021; 800:145834. <u>https://doi.org/10.1016/j.gene.2021.145834</u> PMID:34274483
- Ge L, Zhu J, Liu J, Li L, Zhang J, Cheng J, Li Y, Yang Z, Li S, He J, Zhang X. METTL1 gene polymorphisms synergistically confer hepatoblastoma susceptibility. Discov Oncol. 2022; 13:77. <u>https://doi.org/10.1007/s12672-022-00545-7</u> PMID:<u>35986847</u>
- Li L, Zhuo Z, Yang Z, Zhu J, He X, Yang Z, Zhang J, Xin Y, He J, Zhang T. *HMGA2* Polymorphisms and Hepatoblastoma Susceptibility: A Five-Center Case-Control Study. Pharmgenomics Pers Med. 2020; 13:51–7. <u>https://doi.org/10.2147/PGPM.S241100</u> PMID:<u>32104047</u>
- He S, Zhu J, Xiao Z, Liu J, Zhang J, Li Y, Yang Z, Cheng J, Li S, Li L, He J, Xu D. WDR4 gene polymorphisms increase hepatoblastoma susceptibility in girls. J Cancer. 2022; 13:3342–7.

https://doi.org/10.7150/jca.76255 PMID:36186903

 Luo Z, Li G, Wang M, Zhu J, Yang Z, Li Y, Zhang J, Xin Y, Li S, Li L, Zhuo Z, He J. YTHDF1 rs6090311 A>G polymorphism reduces Hepatoblastoma risk: Evidence from a seven-center case-control study. J Cancer. 2020; 11:5129–34.

https://doi.org/10.7150/jca.46120 PMID:<u>32742460</u>

- Zhuo ZJ, Hua RX, Chen Z, Zhu J, Wang M, Yang Z, Zhang J, Li Y, Li L, Li S, Xin Y, Xia H, He J. WTAP Gene Variants Confer Hepatoblastoma Susceptibility: A Seven-Center Case-Control Study. Mol Ther Oncolytics. 2020; 18:118–25. <u>https://doi.org/10.1016/j.omto.2020.06.007</u> PMID:32671187
- 16. Li J, Zhang H, Wang H. N<sup>1</sup>-methyladenosine modification in cancer biology: Current status and future perspectives. Comput Struct Biotechnol J. 2022; 20:6578–85. <u>https://doi.org/10.1016/j.csbj.2022.11.045</u> PMID:36467585
- 17. Wiener D, Schwartz S. The epitranscriptome beyond m<sup>6</sup>A. Nat Rev Genet. 2021; 22:119–31. <u>https://doi.org/10.1038/s41576-020-00295-8</u> PMID:33188361
- Su Z, Monshaugen I, Wilson B, Wang F, Klungland A, Ougland R, Dutta A. TRMT6/61A-dependent base methylation of tRNA-derived fragments regulates gene-silencing activity and the unfolded protein response in bladder cancer. Nat Commun. 2022; 13:2165.

https://doi.org/10.1038/s41467-022-29790-8 PMID:35444240

- 19. Ye Y, Liu M, Wu F, Ou S, Wang W, Fei J, Xie F, Bai L. TRMT6 promotes hepatocellular carcinoma progression through the PI3K/AKT signaling pathway. Eur J Med Res. 2023; 28:48. <u>https://doi.org/10.1186/s40001-022-00951-1</u> PMID:<u>36707905</u>
- 20. Feng Q, Wang D, Xue T, Lin C, Gao Y, Sun L, Jin Y, Liu D. The role of RNA modification in hepatocellular carcinoma. Front Pharmacol. 2022; 13:984453. <u>https://doi.org/10.3389/fphar.2022.984453</u> PMID:<u>36120301</u>
- Cui L, Ma R, Cai J, Guo C, Chen Z, Yao L, Wang Y, Fan R, Wang X, Shi Y. RNA modifications: importance in immune cell biology and related diseases. Signal Transduct Target Ther. 2022; 7:334. <u>https://doi.org/10.1038/s41392-022-01175-9</u> PMID:<u>36138023</u>
- Wang Y, Wang J, Li X, Xiong X, Wang J, Zhou Z, Zhu X, Gu Y, Dominissini D, He L, Tian Y, Yi C, Fan Z. N<sup>1</sup>-

methyladenosine methylation in tRNA drives liver tumourigenesis by regulating cholesterol metabolism. Nat Commun. 2021; 12:6314. <u>https://doi.org/10.1038/s41467-021-26718-6</u> PMID:34728628

- Chujo T, Suzuki T. Trmt61B is a methyltransferase responsible for 1-methyladenosine at position 58 of human mitochondrial tRNAs. RNA. 2012; 18:2269–76. <u>https://doi.org/10.1261/rna.035600.112</u> PMID:23097428
- Bar-Yaacov D, Frumkin I, Yashiro Y, Chujo T, Ishigami Y, Chemla Y, Blumberg A, Schlesinger O, Bieri P, Greber B, Ban N, Zarivach R, Alfonta L, et al. Mitochondrial 16S rRNA Is Methylated by tRNA Methyltransferase TRMT61B in All Vertebrates. PLoS Biol. 2016; 14:e1002557. https://doi.org/10.1371/journal.pbio.1002557

PMID:27631568

 Couch FJ, Kuchenbaecker KB, Michailidou K, Mendoza-Fandino GA, Nord S, Lilyquist J, Olswold C, Hallberg E, Agata S, Ahsan H, Aittomäki K, Ambrosone C, Andrulis IL, et al. Identification of four novel susceptibility loci for oestrogen receptor negative breast cancer. Nat Commun. 2016; 7:11375. https://doi.org/10.1038/ncomms11375

PMID:27117709

- 26. Ali AT, Idaghdour Y, Hodgkinson A. Analysis of mitochondrial m1A/G RNA modification reveals links to nuclear genetic variants and associated disease processes. Commun Biol. 2020; 3:147. <u>https://doi.org/10.1038/s42003-020-0879-3</u> PMID:<u>32221480</u>
- Martín A, Epifano C, Vilaplana-Marti B, Hernández I, Macías RIR, Martínez-Ramírez Á, Cerezo A, Cabezas-Sainz P, Garranzo-Asensio M, Amarilla-Quintana S, Gómez-Domínguez D, Caleiras E, Camps J, et al. Mitochondrial RNA methyltransferase TRMT61B is a new, potential biomarker and therapeutic target for highly aneuploid cancers. Cell Death Differ. 2023; 30:37–53.

https://doi.org/10.1038/s41418-022-01044-6 PMID:<u>35869285</u>

- Ren H, Zhuo ZJ, Duan F, Li Y, Yang Z, Zhang J, Cheng J, Li S, Li L, Geng J, Zhang Z, He J, Niu H. *ALKBH5* Gene Polymorphisms and Hepatoblastoma Susceptibility in Chinese Children. J Oncol. 2021; 2021:6658480. <u>https://doi.org/10.1155/2021/6658480</u> PMID:33790968
- 29. Fan J, Zhuo Z, Chen G, Niu H, Yang Z, Zhang J, Li Y, Li S, Cheng J, Li L, He J, Wang X. *FTO* gene polymorphisms and hepatoblastoma susceptibility among Chinese

children. Cell Cycle. 2022; 21:1512–8. https://doi.org/10.1080/15384101.2022.2054635 PMID:<u>35311451</u>

30. Chen H, Li Y, Li L, Zhu J, Yang Z, Zhang J, Li S, Xin Y, Xia H, He J. YTHDC1 gene polymorphisms and hepatoblastoma susceptibility in Chinese children: A seven-center case-control study. J Gene Med. 2020; 22:e3249.

https://doi.org/10.1002/jgm.3249 PMID:32729171

- 31. Silva JV, Freitas MJ, Fardilha M. Phosphoprotein phosphatase 1 complexes in spermatogenesis. Curr Mol Pharmacol. 2014; 7:136–46. <u>https://doi.org/10.2174/1874467208666150126154222</u> PMID:<u>25620225</u>
- Mai H, Xie H, Hou J, Chen H, Zhou B, Hou J, Jiang D. A Genetic Variant of PPP1CB Influences Risk of Hepatitis B Virus-Related Hepatocellular Carcinoma in Han Chinese: A Pathway Based Analysis. J Hepatocell Carcinoma. 2021; 8:1055–64. https://doi.org/10.2147/JHC.S321939 PMID:34513747
- 33. Roebuck DJ, Aronson D, Clapuyt P, Czauderna P, de Ville de Goyet J, Gauthier F, Mackinlay G, Maibach R, McHugh K, Olsen OE, Otte JB, Pariente D, Plaschkes J, et al, and International Childrhood Liver Tumor Strategy Group. 2005 PRETEXT: a revised staging system for primary malignant liver tumours of childhood developed by the SIOPEL group. Pediatr Radiol. 2007; 37:123–32. https://doi.org/10.1007/s00247-006-0361-5

PMID:<u>17186233</u>

34. Bian J, Zhuo Z, Zhu J, Yang Z, Jiao Z, Li Y, Cheng J, Zhou H, Li S, Li L, He J, Liu Y. Association between METTL3 gene polymorphisms and neuroblastoma susceptibility: A nine-centre case-control study. J Cell Mol Med. 2020; 24:9280–6. https://doi.org/10.1111/jcmm.15576 PMID:32615646

35. Guan Q, Lin H, Hua W, Lin L, Liu J, Deng L, Zhang J, Cheng J, Yang Z, Li Y, Bian J, Zhou H, Li S, et al. Variant rs8400 enhances ALKBH5 expression through disrupting miR-186 binding and promotes neuroblastoma progression. Chin J Cancer Res. 2023; 35:140–62. https://doi.org/10.21147/j.issn.1000-9604.2023.02.05

PMID:<u>37180836</u>
36. He J, Yuan L, Lin H, Lin A, Chen H, Luo A, Zhuo Z, Liu X. Genetic variants in m<sup>6</sup>A modification core genes are

Genetic variants in m<sup>6</sup>A modification core genes are associated with glioma risk in Chinese children. Mol Ther Oncolytics. 2021; 20:199–208. <u>https://doi.org/10.1016/j.omto.2020.12.013</u> PMID:33665358

- Wacholder S, Chanock S, Garcia-Closas M, El Ghormli L, Rothman N. Assessing the probability that a positive report is false: an approach for molecular epidemiology studies. J Natl Cancer Inst. 2004; 96:434–42. <u>https://doi.org/10.1093/jnci/djh075</u> PMID:<u>15026468</u>
- He J, Zou Y, Liu X, Zhu J, Zhang J, Zhang R, Yang T, Xia H. Association of Common Genetic Variants in PremicroRNAs and Neuroblastoma Susceptibility: A Two-Center Study in Chinese Children. Mol Ther Nucleic Acids. 2018; 11:1–8. <u>https://doi.org/10.1016/j.omtn.2018.01.003</u> PMID:29858046

## SUPPLEMENTARY MATERIALS

# **Supplementary Table**

Variables	Cases (n=313)		Controls	<b>D</b> 9	
Variables	No.	%	No.	%	- P <sup>a</sup>
Age range, month	0.03-149.97		0.004-	0.004-156.00	
$Mean \pm SD$	$23.75\pm25.93$		$25.23 \pm 19.38$		0.251 <sup>b</sup>
<17	168	53.67	642	44.40	
≥17	145	46.33	804	55.60	
Gender					0.983
Female	129	41.21	595	41.15	
Male	184	58.79	851	58.85	
Clinical stages					
Ι	97	30.99	/	/	
II	63	20.13	/	/	
III	64	20.45	/	/	
IV	27	8.63	/	/	
NA	62	19.81	/	/	

Supplementary Table 1. Frequency distribution of selected variables in hepatoblastoma patients and cancer-free controls.

SD, standard deviation; NA, not available.

 $^{a}\text{Two-sided}~\chi^{2}$  test for distributions between hepatoblastoma cases and cancer-free controls.

<sup>b</sup>T-test for age distribution between hepatoblastoma patients and cancerfree controls.