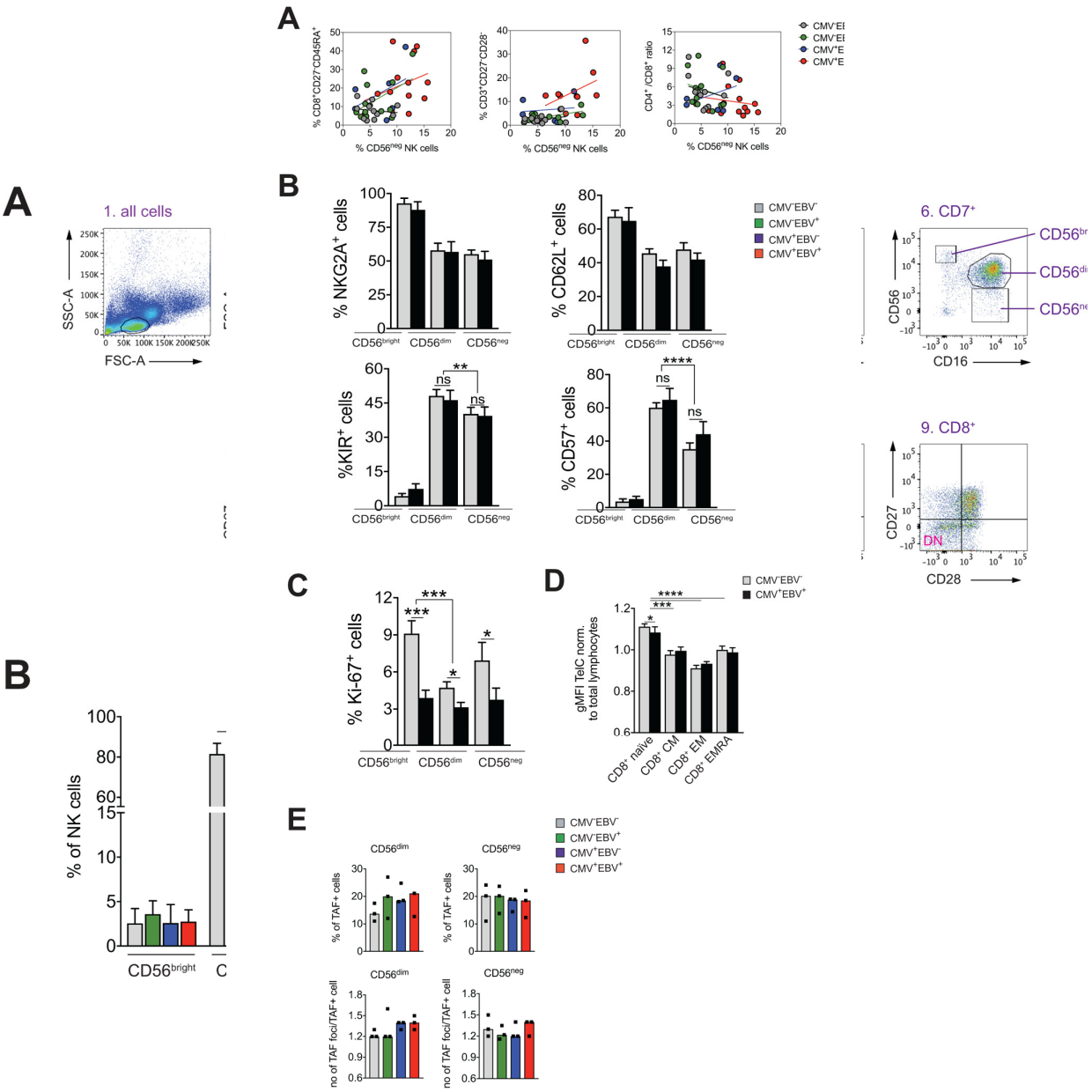

Figure S1. Experimental gating strategy for NK and T cell subsets and frequencies of NK cell subsets within cohort subgroups. (**A**) For all FACS experiments performed on total PBMCs, the following gating strategy was applied: (**1**) Lymphocytes were identified by sideward (SSC) and forward scatter (FSC) parameters. (**2**) Doublet discrimination gating was performed by plotting FSC-area (FSC-A) versus FSC-height (FSC-H) and excluding cells with disproportionate cell size as indicated. (**3**) Dead cells were excluded by staining with a fixable viability dye. Only living cells were used for further analysis. For analysis of NK cells, we first excluded T cells by gating on $CD3^-$ lymphocytes only (**4**), followed by exclusion of $CD7^-$ cells (**5**). These cells were then used to define $CD56^{bright}$, $CD56^{dim}$ and $CD56^{neg}$ NK cells, as indicated in (**6**) T cells were identified by gating on $CD3^+$, $CD56^-$ cells (**4**) followed by CD4 and CD8 gating (**7**). $CD4^+$ and $CD8^+$ T cells were further divided into naïve (N), central memory (CM), effector memory (EM) and terminally-differentiated effector memory (EMRA) subsets, based on their expression of CD27 and CD45RA (**8** and **10**). Double-negative (DN) $CD4^+$ and $CD8^+$ T cells were determined by CD27 and CD28 gating (**9** and **11**). (**B**) Frequencies of $CD56^{bright}$, $CD56^{dim}$ and $CD56^{neg}$ NK cells – as determined by FACS analysis in total PBMCs– are shown in a cohort of HDs >60 years of age stratified as CMV^-EBV^- (n=11/11), CMV^-EBV^+ (n=21/24), CMV^+EBV^- (n=6/6), and CMV^+EBV^+ (n=12/14). * p≤0.05, ** p≤0.005, *** p≤0.005, **** p≤0.0005.